

### GTFCC LABORATORY TRAINING TOOLS

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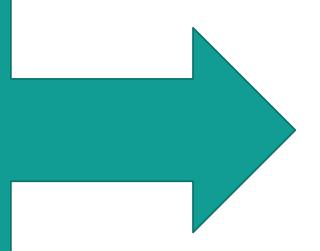
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## BACKGROUND & PREVIOUS DISCUSSIONS

## GTFCC TRAINING SUPPORT

#### **GTFCC Laboratory Training Toolkit**

- Develop standardized material (presentations, training plans, checklists, etc.)
- Toolkit available via GTFCC website
- Online training courses (OpenWHO)



Laboratories and stakeholders can access and use the materials as needed + use for GTFCC trainings

## BENEFITS OF LABORATORY TRAINING

- Build increased capacity for lab confirmation
- Improve ability to inform decision making
- Promote accuracy and consistency for testing and reporting results
- Support the development of staff / increase competence
- Improve staff preparedness and response
- Improve quality of the test system
- Better confidence in results

# WHAT ARE THE KEY TRAINING TOPICS?

- Cholera basics (e.g., disease, pathogen, response, etc.)
  - Context can be broad or very specific
- Detail will depend on the audience
- Specimen collection, preservation and transport
- Adaptive testing strategy
- Rapid Diagnostic Tests (RDTs)
- Primary isolation of Vibrio cholerae from stool specimens
- Strain conditioning for shipment and storage (short-term and long-term)
- •Identification of toxigenic Vibrio cholerae O1 / O139
- Culture-based methods (e.g., agglutination in VC-specific antisera)
- Molecular methods (e.g., PCR)
- Antimicrobial susceptibility testing (AST)
- Data management and reporting laboratory results

## ADDITIONAL ASPECTS OF THE TRAININGS

■Who should be trained?

Field staff, lab staff, and higher-level decision makers

☐ How to structure the training?

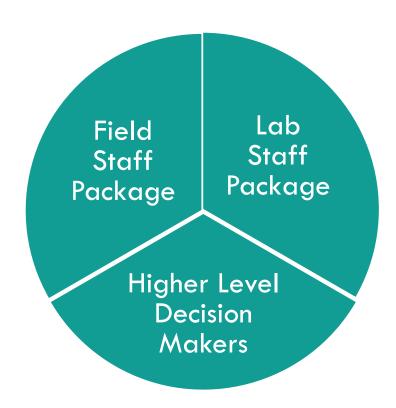
Stepwise approach for the delivery covering the topics previously shown

□ Validating and measuring the effectiveness of the training

## PROGRESS IN 2023-2024

## STANDARDIZED TRAINING MATERIALS

☐ 3 different training packages targeting the various arms involved in cholera testing and response.



## FIELD STAFF PACKAGE: INTRODUCTION TO TESTING FOR CHOLERA

#### ■Comprises of 3 modules:

- ☐ Introduction to Cholera
- Sample Collection, Preparation, and Transport for Cholera
- □ Cholera Rapid Diagnostic Tests (RDTs)

Was sent out for review to all the LWG members!

#### ☐ Target audience:

- Rapid response teams
- □Surveillance field teams
- Nurses
- Medical doctors
- Other primary healthcare workers
- Laboratory personnel (including from reference laboratories).

## LAB STAFF PACKAGE: BASIC LABORATORY METHODS FOR CHOLERA CONFIRMATION

- □ Comprises of 8 modules:
  - ☐ Introduction to Cholera
  - ☐ Testing Algorithms
  - ■Sample Receipt
  - Media Preparation
  - □ Culture confirmation
  - Antimicrobial Susceptibility
  - Reporting of Results
  - ■Additional Tests

- ☐ Target audience:
- Laboratory directors
- Laboratory personnel involved in cholera testing



# HIGHER LEVEL DECISION MAKERS PACKAGE: TESTING STRATEGY AND CONSIDERATION FOR LABORATORY PREPAREDNESS AND RESPONSE FOR CHOLERA

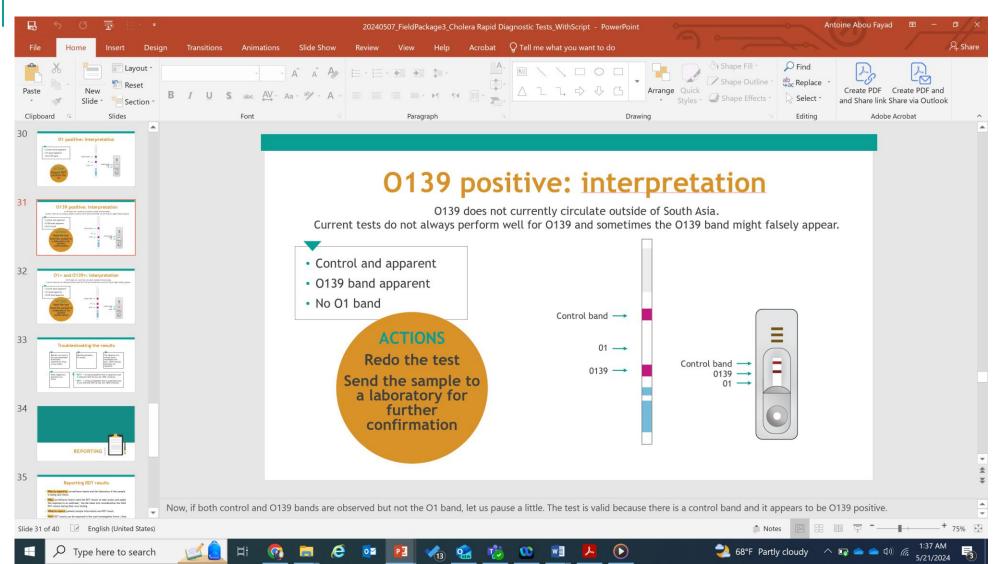
- ■Comprises of 3 modules:
- ☐ Testing Strategy
- ☐ Preparedness for Cholera
- Response to a Cholera Outbreak

Will be sent out for review to all the LWG members!

#### ☐ Target audience:

- Rapid response team leads
- ■Surveillance field team leads
- ☐ Ministry of Public Health staff.
- Laboratory directors.

## SNAPSHOTS FROM THE TRAINING MATERIAL



## TRAINING OF TRAINERS (TOT)

Further support building capacity for cholera surveillance

Improve the response and the understanding of the in-country burden of cholera.

#### **Objectives of the ToT:**

- 1- Provide laboratories with trainers for cholera diagnostics  $\rightarrow$  cascade trainings across countries.
- 2- Directly improve the capacity for testing and confirming cholera in country.
- 3- Improved cholera surveillance.

Funded by CDC and IMST to perform emergency ToT in high priority countries

## TOPICS AND CONTENT OF TOT

| Topics  | Setup required                                       |  |
|---|--|--|
| Introduction to cholera   | Interactive lecture                                  |  |
| Stool sample collection and sample preparation for transport  | Interactive lecture and practical laboratory session |  |
| Use of rapid diagnostic tests   | Interactive lecture and practical laboratory session |  |
| Culture methods for cholera with focus on GTFCC recommended methods (culture/oxidase/seroagglutination) | Interactive lecture and practical laboratory session |  |
| Antimicrobial susceptibility testing  | Interactive lecture and practical laboratory session |  |
| Other laboratory methods for cholera  | Interactive lecture                                  |  |
| Reporting of results  | Interactive lecture                                  |  |
| Strategic testing for cholera   | Interactive lecture                                  |  |
| How to manage and plan a training   | Interactive lecture                                  |  |

## TOT AGENDA

- □9 days long
  - First 6 days will be working with the "trainers to be"
  - Last 3 days of the training will be centered around setting up cascade trainings by the new trainers and attending those trainings



#### Cholera laboratory diagnostics training of trainers

The Republic of XXX

**▲** 4-14 July, 2024

#### **Training Agenda**

Day 1

| Time   | Activity Title   | Required Setup |  |
|--|--|----------------|--|
| 8:00-8:30  | <ul> <li>Opening Remarks</li> </ul>                      | Classroom      |  |
|  | <ul> <li>Goals for the workshop</li> </ul>               |                |  |
|  | <ul> <li>Introduction of participants</li> </ul>         |                |  |
|  | <ul> <li>Facility information</li> </ul>                 |                |  |
|  | <ul> <li>Group photo</li> </ul>                          |                |  |
|  | <ul> <li>Overview of the program</li> </ul>              |                |  |
| 8:30-9:00  | Pre-training assessment                                  | Classroom      |  |
| 9:00-10:00   | Introduction to cholera                                  | Classroom      |  |
| 10:00-10:30  | Biosafety and Biosecurity                                | Classroom      |  |
| 10:30-10:45  | Break  |                |  |
| 10:45-12.00  | Sample collection and transport                          | Classroom      |  |
| 12:00-13:00  | Culture - Alkaline Peptone Water (APW) - Media selection | Classroom      |  |
| 13:00-14:00  | Break for lunch  |                |  |
| 14:00-15:00  | Media preparation  | Laboratory     |  |
| 15:00-16:00 Rapid Diagnostic Tests (RDT) - sample qualitesting plan-list of supplies |  | - Laboratory   |  |
|  | Pouring media and plates - RDTs                          | Laboratory     |  |
| 16:00-17:30  | · · · · · · · · · · · · · · · · · ·                      | ,              |  |

#### Day 2

| Time        | Activity Title  | Required Setup |
|-------------|---|----------------|
| 8:00- 8:30  | Recap day 1   | Classroom      |
| 8:30-10:00  | Culture - Direct plating - Inoculation of APW   | Laboratory     |
| 10:00-10:15 | Break   |                |
| 10:45-12:00 | Culture - Isolating a single colony - Oxidase<br>testing – Agglutination - Gram-staining -<br>Analytical Profile Index (API) - Other techniques | Classroom      |
| 12:00-13:00 | Break for Lunch   |                |
| 10:45-12.00 | Sample collection and transport   | Classroom      |
| 13:00-14:00 | AST   | Classroom      |
| 14:00-17:00 | RDT (repeat) and culture inoculations   | Laboratory     |

#### Day 3

| Time        | Activity Title  | Required Setup |
|-------------|---|----------------|
| 8:00- 8:30  | Recap day 2   | Classroom      |
| 8:30-10:30  | Culture - Isolating a single colony -<br>Morphological analysis of the colonies | Laboratory     |
| 10:30-10:45 | Break   |                |
| 10:45-13:00 | Culture - Oxidase testing – Agglutination - API                                 | Laboratory     |
| 13:00-14:00 | Break for Lunch   |                |
| 14:00-17.00 | AST   | Laboratory     |

#### Day 4

| Time         | Activity Title                                  | Required Setup |
|--------------|---|----------------|
| 8:00- 8:30   | Recap day 3                                     | Classroom      |
| 8:30-10:30   | AST analysis                                    | Laboratory     |
| 10:30-10:45  | Break   |                |
| 10:45-13:00  | Culture - Oxidase testing – Agglutination – API | Laboratory     |
| 13:00-14:00  | Break for Lunch                                 |                |
| 14:00-15:30  | Reporting and data analysis (part 1)            | Classroom      |
| 15:30-16:00  | Break   |                |
| 16:00-17 :00 | Reporting and data analysis (part 2)            | Classroom      |
|              |   |                |

### TOT VALIDATION & EVALUATION

#### **ToT Assessment Test**

- 1- Sample collection for culture confirmation should happen after antibiotic treatment: True or False
- 2- Stool sample in a <u>fecal</u> collection cup can be tested by culture in the lab after---- from collection:

3- Swabs in Cary Blair should be transported at 4 °C True or False

4- RDTs cannot detect cholera toxins: True or False

- A positive RDT alone is sufficient to declare the presence of O139 positive sample
   True or False
- 6- If your RDT indicates the presence of O1 but the control line is not visible, then the result of the RDT is considered to be positive. True or False
- 7- Alkaline peptone can be streaked on selective and nonselective culture media after more than 6 hours of incubation.
- 8- Any yellow colony is growing on TCBS agar means that this colony is Vibrio cholerae.
  True or False
- Performing agglutination of Vibrio cholerae using specific antisera allows to identify serogroup and toxigenicity.

True or False

10- What are the minimum antibiotics recommended for Vibrio cholerae sensitivity testing for surveillance and treatment of cholera (please select all correct answers).

a. Ciprofloxacin f. Nalidixic acid b. Amikacin g. Azithromycin c. Meropenem h. Gentamycin d. Colistin j. Minocycline

e. Tetracycline.

11- Using PCR, we can identify Vibrio cholerae (genus and species), serogroup, serotype, confirm the presence of the toxin gene and obtain antimicrobial sensitivity data.

True or False

#### **ToT Participants Feedback Questionnaire**

|          |                      |   |                             |                            | provide them to the instructor or writ |  |  |
|----------|----------------------|---|-----------------------------|----------------------------|--|--|--|
| hen      |                      | Scale: 5 = Strongly Agr   | ee, 4 = Agree, 3 = Neutral, | 2 = Disagree, 1 = Strongly | y Disagree).                           |  |  |
|          |                      |   |                             |                            |  |  |  |
| tro      | ngly Agree           | Agree   | Neutral                     | Disagree                   | Strongly Disagree                      |  |  |
| <u>.</u> | The quality and co   | ontent of the training ma   | terials and presentations   | met my expectations.       |  |  |  |
| tro      | ngly Agree           | Agree   | Neutral                     | Disagree                   | Strongly Disagree                      |  |  |
|          |                      |   |                             |                            |  |  |  |
| 3.       | The instructor or    | e instructor or teacher was aware/familiar with the topic they discussed and its relevance to my country. |                             |                            |  |  |  |
| tro      | ngly Agree           | Agree   | Neutral                     | Disagree                   | Strongly Disagree                      |  |  |
| ŧ.       | The ability clarity  | and completeness of th  | e instructor were adequa    | te when responding to tr   | ainee questions                        |  |  |
|          | The ability, clarity | and completeness of the   | instructor were adequa      | te when responding to the  | antee questions.                       |  |  |
| itro     | ngly Agree           | Agree   | Neutral                     | Disagree                   | Strongly Disagree                      |  |  |
| 5.       | The training conte   | ent provided enough con   | text and went into enoug    | h detail.                  |  |  |  |
| tro      | l<br>ngly Agree      | Agree   | Neutral                     | Disagree                   | Strongly Disagree                      |  |  |
| 5.       | The trainer was e    | ngaging and supportive.   |                             |                            |  |  |  |
|          |                      |   |                             | T                          |  |  |  |
| tro      | ngly Agree           | Agree   | Neutral                     | Disagree                   | Strongly Disagree                      |  |  |
| 7.       | The group discuss    | ions adequately covered   | issues of concern releva    | nt to my country.          |  |  |  |
|          | ngly Agree           | Agree   | Neutral                     | Disagree                   | Strongly Disagree                      |  |  |
| tre      | P.1 - (B) ==         | - Agree   | I VEGUTAL                   | Disablee                   | Strongly bisagree                      |  |  |
| Stron    |                      |   |                             |                            |  |  |  |

## PILOT TRAININGS

These trainings were piloted in South Sudan, Somalia, and Union of Comoros so far.





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## FUTURE PERSPECTIVES

- ☐ Finalize all the toolkit
- ☐ Publish the toolkit on GTFCC website as they are completed
- ☐ Transform the toolkit into OpenWHO courses
- ☐ Continue setting up more ToTs, including regional level ToTs!



## THANK YOU