**Specimen Packaging and Domestic Transportation for Laboratory Confirmation of Vibrio cholerae O1/O139**

### FAECAL SPECIMENS CONDITIONING: 4 possible options

<table>
<thead>
<tr>
<th>Faecal Specimen in Stool Container</th>
<th>APW (alkaline peptone water)</th>
<th>Wet and Dry Filter Paper (WFP/DFP)</th>
<th>Cary Blair medium, Faecal Sample or Rectal Swab</th>
</tr>
</thead>
<tbody>
<tr>
<td>Keep in initial stool container.</td>
<td>Transfer faecal material from initial container into APW tube.  <strong>NOTE:</strong> The faecal material should not exceed 10% of the volume of the APW enrichment.</td>
<td>Dip filter disk into watery faecal material with single-use device (forceps, needle), transfer into tube, add 2 to 3 drops of saline, close tube. <strong>DRY FILTER PAPER (DFP)</strong> Deposit one drop of watery stool onto filter paper. Air dry paper before placing into individual pouch with desiccant.</td>
<td>For faecal samples: dip swab in liquid stool and transfer into Cary Blair medium. Rectal swab: Place swab directly into Cary Blair. No further manipulation is required.</td>
</tr>
</tbody>
</table>

Compatibility with testing methods (either directly from sample or after incubation steps in APW for those marked with *).

- RDT, culture, molecular analysis
- RDT, culture, molecular analysis
- WFP: culture, molecular analysis, RDT*
- WFP: culture, molecular analysis, DFP: molecular analysis
- Culture, molecular analysis* and RDT*

### MATERIAL REQUIRED

- Stool container (plastic, screw cap, 30ml, without disinfectant)
- APW, tubes with screw cap, transfer pipettes or swabs
- WFP: Filter paper discs (6mm Ø, non-sterile), saline solution, forceps or needle, 2ml tube (screw cap)
- DFP: Whatman cards (903 protein saver, FTA Elute Micro Cards), disposable transfer pipettes, individual pouches, desiccant
- Cary Blair (semi-solid, bottle/tube), swab (sterile, cotton/polyester)

Paraffilm or sealing tape to seal packages and prevent leakage (not required for dry filter paper).

### CONSERVATION

Ambient temperature (ideally 22-25°C). Do not refrigerate. Keep stool container out of direct sunlight.

- 2 hours max. If delay >2h, use Cary Blair.
- Less than 24 hours
- WFP: ideally less than 15 days DFP: no limitation
- Follow manufacturer’s instructions, on average 7 days

### DOMESTIC TRANSPORTATION (national shipment, by road)

<table>
<thead>
<tr>
<th>Primary Containers</th>
<th>Absorbent Materials</th>
<th>Secondary Container</th>
<th>Tertiary Containers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Primary containers are individually wrapped, and absorbent material placed between the primary container(s) and the secondary containers.</td>
<td>Cellulose Wadding Cotton Balls Paper Towels</td>
<td>Sealed Polystyrene Container (1-inch-think minimum) Sealed Plastic Bag Plastic Canister Screw-Cap Can</td>
<td>Rigid cooler without ice-pack Compressed fiberglass Biological Substance Category B</td>
</tr>
</tbody>
</table>

Samples are categorized “biological substances” category B. The use of triple packaging with UN3373 labels are required, alternatives are shown on the left.

Samples must travel with corresponding documentation (lab request form and/or line list). Include any results that may have already been performed, such as RDT results. Do not write the name of the organism on the outside of the package, only on the paperwork inside the box where appropriate.

**IMPORTANT:** indicate complete address and phone number for sender and recipient. Inform recipient laboratory about upcoming arrival of samples.

**TRANSPORT AT AMBIENT TEMPERATURE**

GTFC, April 2019

1 Annex 2B: Case-based laboratory reporting form from: https://www.afro.who.int/sites/default/files/2017-06/IDSR-Technical-Guidelines_Final_2010_0.pdf